

FORM PTO-1390 (Modified) (REV 11-2000)		U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE		ATTORNEY'S DOCKET NUMBER 8830-23
TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371				U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR 10/049706
INTERNATIONAL APPLICATION NO. PCT/GB00/03223	INTERNATIONAL FILING DATE August 18, 2000	PRIORITY DATE CLAIMED August 19, 1999		
TITLE OF INVENTION Trehalose Producing Prokaryotic Cells As Vaccines				
APPLICANT(S) FOR DO/EO/US Camilo Anthony Leo Selwyn Colaco				
Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:				
<ol style="list-style-type: none"> 1. <input checked="" type="checkbox"/> This is a FIRST submission of items concerning a filing under 35 U.S.C. 371. 2. <input type="checkbox"/> This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371. 3. <input checked="" type="checkbox"/> This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (24) indicated below. 4. <input checked="" type="checkbox"/> The US has been elected by the expiration of 19 months from the priority date (Article 31). 5. <input checked="" type="checkbox"/> A copy of the International Application as filed (35 U.S.C. 371 (c) (2)) <ol style="list-style-type: none"> a. <input type="checkbox"/> is attached hereto (required only if not communicated by the International Bureau). b. <input checked="" type="checkbox"/> has been communicated by the International Bureau. c. <input type="checkbox"/> is not required, as the application was filed in the United States Receiving Office (RO/US). 6. <input type="checkbox"/> An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)). <ol style="list-style-type: none"> a. <input type="checkbox"/> is attached hereto. b. <input type="checkbox"/> has been previously submitted under 35 U.S.C. 154(d)(4). 7. <input checked="" type="checkbox"/> Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3)) <ol style="list-style-type: none"> a. <input type="checkbox"/> are attached hereto (required only if not communicated by the International Bureau). b. <input type="checkbox"/> have been communicated by the International Bureau. c. <input type="checkbox"/> have not been made; however, the time limit for making such amendments has NOT expired. d. <input checked="" type="checkbox"/> have not been made and will not be made. 8. <input type="checkbox"/> An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)). 9. <input type="checkbox"/> An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)). 10. <input type="checkbox"/> An English language translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)). 11. <input checked="" type="checkbox"/> A copy of the International Preliminary Examination Report (PCT/IPEA/409). 12. <input checked="" type="checkbox"/> A copy of the International Search Report (PCT/ISA/210). <p>Items 13 to 20 below concern document(s) or information included:</p> <ol style="list-style-type: none"> 13. <input checked="" type="checkbox"/> An Information Disclosure Statement under 37 CFR 1.97 and 1.98. 14. <input type="checkbox"/> An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included. 15. <input checked="" type="checkbox"/> A FIRST preliminary amendment. 16. <input type="checkbox"/> A SECOND or SUBSEQUENT preliminary amendment. 17. <input type="checkbox"/> A substitute specification. 18. <input type="checkbox"/> A change of power of attorney and/or address letter. 19. <input type="checkbox"/> A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821 - 1.825. 20. <input type="checkbox"/> A second copy of the published international application under 35 U.S.C. 154(d)(4). 21. <input type="checkbox"/> A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4). 22. <input checked="" type="checkbox"/> Certificate of Mailing by Express Mail 23. <input checked="" type="checkbox"/> Other items or information: <p>US Express Mail No. EL 931090059 US Courtesy Copy of PCT/GB00/03223 Publication Unexecuted Declaration and Power of Attorney</p>				

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR 10/049706	INTERNATIONAL APPLICATION NO. PCT/GB00/03223	ATTORNEY'S DOCKET NUMBER 8830-23																
24. The following fees are submitted:		CALCULATIONS PTO USE ONLY																
BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)) :																		
<input type="checkbox"/> Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO \$1040.00 <input checked="" type="checkbox"/> International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$890.00 <input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$740.00 <input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$710.00 <input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) \$100.00																		
ENTER APPROPRIATE BASIC FEE AMOUNT =		\$890.00																
Surcharge of \$130.00 for furnishing the oath or declaration later than months from the earliest claimed priority date (37 CFR 1.492 (e)).		<input type="checkbox"/> 20 <input type="checkbox"/> 30 \$0.00																
<table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="width: 25%;">CLAIMS</th> <th style="width: 25%;">NUMBER FILED</th> <th style="width: 25%;">NUMBER EXTRA</th> <th style="width: 25%;">RATE</th> </tr> </thead> <tbody> <tr> <td>Total claims</td> <td>21 - 20 =</td> <td>1</td> <td>x \$18.00</td> </tr> <tr> <td>Independent claims</td> <td>3 - 3 =</td> <td>0</td> <td>x \$84.00</td> </tr> <tr> <td colspan="2">Multiple Dependent Claims (check if applicable)</td> <td></td> <td style="text-align: center;">□</td> </tr> </tbody> </table>		CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE	Total claims	21 - 20 =	1	x \$18.00	Independent claims	3 - 3 =	0	x \$84.00	Multiple Dependent Claims (check if applicable)			□	\$18.00
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE															
Total claims	21 - 20 =	1	x \$18.00															
Independent claims	3 - 3 =	0	x \$84.00															
Multiple Dependent Claims (check if applicable)			□															
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TOTAL OF ABOVE CALCULATIONS =		\$908.00																
<input checked="" type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27). The fees indicated above are reduced by 1/2.		\$454.00																
SUBTOTAL =		\$454.00																
Processing fee of \$130.00 for furnishing the English translation later than months from the earliest claimed priority date (37 CFR 1.492 (f)).		<input type="checkbox"/> 20 <input type="checkbox"/> 30 + \$0.00																
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TOTAL NATIONAL FEE =		\$454.00																
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable).		<input type="checkbox"/> \$0.00																
TOTAL FEES ENCLOSED =		\$454.00																
		Amount to be: refunded																
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a. <input checked="" type="checkbox"/> A check in the amount of \$454.00 to cover the above fees is enclosed. b. <input type="checkbox"/> Please charge my Deposit Account No. _____ in the amount of _____ to cover the above fees. A duplicate copy of this sheet is enclosed. c. <input checked="" type="checkbox"/> The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 50-0573 A duplicate copy of this sheet is enclosed. d. <input type="checkbox"/> Fees are to be charged to a credit card. WARNING: Information on this form may become public. Credit card information should not be included on this form. Provide credit card information and authorization on PTO-2038.																		
NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.																		
SEND ALL CORRESPONDENCE TO: <div style="border: 1px solid black; padding: 5px; margin-bottom: 10px;"> DANIEL A. MONACO Drinker Biddle & Reath LLP One Logan Square 18th and Cherry Streets Philadelphia, Pennsylvania 19103-6996 (215) 988-3312 (215) 988-2757 Fax </div> <div style="text-align: right; margin-bottom: 10px;">  DANIEL A. MONACO </div> <div style="text-align: right; margin-bottom: 10px;"> NAME </div> <div style="text-align: right; margin-bottom: 10px;"> 30,480 </div> <div style="text-align: right; margin-bottom: 10px;"> REGISTRATION NUMBER </div> <div style="text-align: right; margin-bottom: 10px;"> February 14, 2002 </div> <div style="text-align: right; margin-bottom: 10px;"> DATE </div>																		

10/049706
JC11 Rec'd PCT/PTO 14 FEB 2002

PATENT
Attorney Docket No.: 8830-23

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re: Patent application of
Camilo Anthony Leo Selwyn Colaco : Group Art Unit:
Serial No.: not yet assigned
(International application: PCT/GB00/03223)
Filed: not yet assigned : Examiner:
(International filing date: August 18, 2000)
For: TREHALOSE PRODUCING PROKARYOTIC
CELLS AS VACCINES

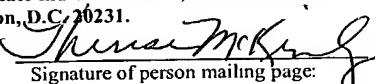
PRELIMINARY AMENDMENT

Commissioner for Patents
Washington, D.C. 20231

Kindly amend the above-identified patent application, without prejudice, in advance of
calculation of the filing fee.

In the Specification:

Insert the abstract attached hereto on a separate page.

CERTIFICATE OF MAILING UNDER 37 C.F.R. 1.10	
EXPRESS MAIL Mailing Label Number: EL 931090059 US	
Date of Deposit: February 14, 2002	
I hereby certify that this correspondence, along with any paper referred to as being attached or enclosed, and/or fee, is being deposited with the United States Postal Service, "EXPRESS MAIL-POST OFFICE TO ADDRESSEE" service under 37 C.F.R. 1.10, on the date indicated above, and addressed to: Commissioner for Patents, Washington, D.C. 20231.	
 Signature of person mailing page: Therese McKinley	
Type or print name of person	

In the Claims:

Cancel claim 19.

Rewrite claims 1-18 and 20-22 to read as follows. A mark-up of the amended claims is contained in Appendix A.

1. (amended) A method for producing a vaccine composition containing an immunogenic determinant as the active ingredient, the method comprising the steps of:
 - a. treating procaryotic cells under conditions such that an increase of the concentration of trehalose within the procaryotic cells is induced;
 - b. using the induced cells containing trehalose as the immunogenic determinant in the production of a vaccine composition.
2. (amended) The method as claimed in claim 1, wherein the treatment of the procaryotic cells is carried out to achieve a concentration of trehalose within the cells of at least 10 mM.
3. (amended) The method as claimed in claim 1, wherein the increase in concentration of trehalose is achieved by synthesis of trehalose within the cell.
4. (amended) The method as claimed in claim 1, wherein the condition causing the increase of trehalose concentration within the cells is heat, osmotic shock, suppression of degradation of trehalose, or genetically engineered constitutive synthesis of trehalose within the cells.
5. (amended) The method as claimed in claim 1, wherein the induced cells containing the trehalose are dried prior to their use in the production of the vaccine composition.
6. (amended) The method as claimed in claim 5, wherein the cells are dried in the absence of added extra-cellular carbohydrate glassy stabilising matrix.
7. (amended) The method as claimed in claim 1, wherein the procaryotic cells are bacteria, protozoa or fungi.

8. (amended) The method as claimed in claim 1, wherein the procaryotic cells are treated by cultivating them in a medium containing one or more solutes and having an osmolarity of at least 350 mOsmoles.

9. (amended) The method as claimed in claim 8, wherein the solute is selected from [a] the group consisting of sodium, potassium, calcium and ammonium salts, and combinations thereof.

10. (amended) The method as claimed in claim 1, wherein the procaryotic cell has been modified so as to synthesise trehalose.

11. (amended) The method as claimed in claim 1, wherein the treatment of the cells is carried out to achieve a concentration of trehalose within the cells of at least 100mM.

12. (amended) The method as claimed in claim 1, wherein the procaryotic cells containing the induced trehalose are killed prior to use in the vaccine composition.

13. (amended) The method as claimed in claim 1, wherein the treatment of the procaryotic cells is carried out in vitro.

14. (amended) A vaccine composition comprising an immunogenic determinant, wherein the immunogenic determinant includes a procaryotic cell or cell residue which contains at least 10mM of trehalose within the cell.

15. (amended) A vaccine composition comprising an immunogenic determinant produced by the method of claim 1.

16. (amended) The vaccine composition as claimed in claim 14, comprising an adjuvant for the immunogenic determinant.

17. (amended) The vaccine composition as claimed in claim 14, comprising an aqueous carrier.

18. (amended) A vaccine composition as claimed in claim 14, wherein the induced cells containing trehalose have been dried in the presence of a non-reducing carbohydrate to provide a storage stable but viable immunogenic determinant for storage prior to use in [a] the vaccine composition.

20. (amended) A method for treating an animal with a vaccine, comprising administering to said animal a pharmaceutically effective amount of a vaccine composition as claimed in claim 1 to elicit an immune response in the animal.

21. (amended) The method as claimed in claim 20, wherein the vaccine composition is administered by injection.

22. (amended) A prokaryotic cell which has had its genetic structure modified so as to remove or inhibit that portion of the genetic structure which inhibits or restricts the synthesis of trehalose by the cell, whereby the cell constitutively synthesises trehalose within the cell as it grows.

Remarks

Claims 1-18 and 20-22 are pending in the application. The claims were amended in the international phase, as set forth in the Annex to the International Preliminary Examination Report. The claims have been further rewritten herein to reduce dependencies and conform to US practice.

Respectfully submitted,

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APPENDIX A: Mark-up of Amended Claims

1. (amended) A method for producing a vaccine composition containing an immunogenic determinant as the active ingredient, [characterised in that] the method comprising [comprises] the steps of:
 - a. treating procaryotic cells under conditions such that an increase of the concentration of trehalose within the procaryotic cells is induced;
 - b. using the induced cells containing trehalose as the immunogenic determinant in the production of a vaccine composition.
2. (amended) [A] The method as claimed in claim 1, [characterised in that] wherein the treatment of the procaryotic cells is carried out to achieve a concentration of trehalose within the cells of at least 10 mM.
3. (amended) [A] The method as claimed in claim 1, wherein [either of claims 1 or 2, characterised in that] the increase in concentration of trehalose is achieved by synthesis of trehalose within the cell.
4. (amended) [A] The method as claimed in claim 1, wherein [any one of the preceding claims, characterised in that] the condition causing the increase of trehalose concentration within the cells is heat, osmotic shock, suppression of degradation of trehalose, or genetically engineered constitutive synthesis of trehalose within the cells.
5. (amended) [A] The method as claimed in claim 1, wherein [any one of the preceding claims, characterised in that] the induced cells containing the trehalose are dried prior to their use in the production of the vaccine composition.
6. (amended) [A] The method as claimed in claim 5, [characterised in that] wherein the cells are dried in the absence of added extra-cellular carbohydrate glassy stabilising matrix.

APPENDIX A: Mark-up of Amended Claims

7. (amended) [A] The method as claimed in claim 1, wherein [any one of the preceding claims, characterised in that] the procaryotic cells are bacteria, protozoa or fungi.
8. (amended) [A] The method as claimed in claim 1, wherein [any one of the preceding claims, characterised in that] the procaryotic cells are treated by cultivating them in a medium containing one or more solutes and having an osmolarity of at least 350 mOsmoles.
9. (amended) [A] The method as claimed in claim 8, wherein [characterised in that] the solute is selected from [a] the group consisting of sodium, potassium, calcium and [/ or] ammonium [salt] salts, and combinations thereof.
10. (amended) [A] The method as claimed in claim 1, wherein [characterised in that] the procaryotic cell has been modified so as to synthesise trehalose.
11. (amended) [A] The method as claimed in claim 1, wherein [characterised in that] the treatment of the cells is carried out to achieve a concentration of trehalose within the cells of at least 100mM.
12. (amended) [A] The method as claimed in claim 1, wherein [any one of the preceding claims, characterised in that] the procaryotic cells containing the induced trehalose are killed prior to use in the vaccine composition.
13. (amended) [A] The method as claimed in claim 1, wherein [any one of the preceding claims, characterised in that] the treatment of the procaryotic cells is carried out in vitro.
14. (amended) A vaccine composition comprising an immunogenic determinant, [characterised in that] wherein the immunogenic determinant includes a procaryotic cell or cell residue which contains at least 10mM of trehalose within the cell.

APPENDIX A: Mark-up of Amended Claims

15. (amended) A vaccine composition comprising [characterised in that it contains] an immunogenic determinant produced by the method of claim 1 [any of claims 1 to 13].
16. (amended) [A] The vaccine composition as claimed in claim 14, comprising [either of claims 14 or 15, characterised in that it contains] an adjuvant for the immunogenic determinant.
17. (amended) [A] The vaccine composition as claimed in claim 14, comprising [any one of claims 14 to 16, characterised in that it contains] an aqueous carrier.
18. (amended) A vaccine composition as claimed in claim 14, wherein [any one of claims 14 to 17, characterised in that] the induced cells containing trehalose [are] have been dried in the presence of a non-reducing carbohydrate to provide a storage stable but viable immunogenic determinant for storage prior to use in [a] the vaccine composition.
20. (amended) A method for treating an animal with a vaccine, comprising administering to said animal [characterised in that] a pharmaceutically effective amount of a vaccine composition as claimed in claim 1 [any one of claims 14 to 18 is administered to the animal] to elicit an immune response in the animal.
21. (amended) [A] The method as claimed in claim 20, wherein [characterised in that] the vaccine composition is administered by injection.
22. (amended) A prokaryotic cell which has had its genetic structure modified so as to remove or inhibit that portion of the genetic structure which inhibits or restricts the synthesis of trehalose by the cell, whereby the cell constitutively synthesises trehalose within the cell as it grows.

TREHALOSE PRODUCING PROKARYOTIC CELLS AS VACCINES

Abstract of the Disclosure

The present invention relates to methods for using prokaryotic cells which have been modified or induced to synthesize trehalose as vaccines and to vaccine compositions obtained thereby.

TITLE: TREHALOSE PRODUCING CELLS AS VACCINES

This invention relates to the field of vaccines. More specifically, it relates to methods of producing vaccines 5 of trehalose containing prokaryotic cells and the compositions obtained thereby.

BACKGROUND TO THE INVENTION:

10 Prokaryotic cells, particularly bacteria, are widely and increasingly used in medical, agricultural and industrial applications. Agricultural, or environmental, applications include biopesticides and bioremediation. Medical applications include use of bacteria in vaccines 15 as well as for production of pharmaceutical products for other treatments.

For the prokaryotic cells to be used effectively, both in terms of desired results and cost, the cells must be able 20 to be stored for significant periods of time whilst preserving their viability. The term viability is used herein to denote that the cells manifest the features of a functioning living organism, such as metabolism and cell division.

25 Methods for preserving live prokaryotic cells suffer from several serious drawbacks, such as being energy-intensive and requiring cold storage. Thus, freeze-drying is often used for preservation and storage of prokaryotic cells. 30 However, it has the undesirable characteristic of significantly reducing viability of the cells, as well as being time- and energy-intensive and thus expensive.

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PCT GB97/03375 describes a process of stabilising prokaryotic cells by the induction of trehalose synthesis and the drying of the resulting cells in a glassy 5 carbohydrate matrix. This process gives stabilised cells that can be stored at ambient temperatures without loss of viability. Trehalose, (α -D-glucopyranosyl- α -D-glucopyranoside), is a naturally occurring, non-reducing disaccharide which was initially found to be associated 10 with the prevention of desiccation damage in certain plants and animals which can dry out without damage and can revive when re-hydrated. Trehalose has been shown to be useful in preventing denaturation of proteins, viruses and foodstuffs during desiccation, see U.S. Patents Nos. 15 4,891,319; 5,149,653; 5,026,566; Colaco et al. (1992) *Bio/Tech.* 10:1007-1011.

Trehalose synthesis in prokaryotic cells is induced by a number of methods including osmotic shock which induces 20 the endogenous production of trehalose, Welsh et al. (1991) *J. Gen. Microbiol.* 137:745-750.

PCT application No. GB94/01556 describes a process of improving the viability of bacterial dried cells by the 25 induction of trehalose synthesis by nutrient limitation, heat shock or osmoadaptation. PCT application No. GB97/03375 describes a method for the preservation of prokaryotic cells by the drying of cells in a carbohydrate matrix after the induction of trehalose synthesis. The 30 latter invention provides compositions of dried cells that can be stored at ambient temperatures and thus enable a number of industrial applications.

Surprisingly, we have now found that the dried, stabilised procaryotic cells produced by the above methods, are more immunogenic than fresh live cells and hence have
5 particular value as the immunogenic determinant active component in vaccine compositions. Furthermore, we have also found that this increased immunogenicity of the stabilised procaryotic cells is not dependent on the drying process considered essential in the above
10 stabilisation processes, but results from the induction of trehalose synthesis. Although more pronounced with dried cells, this increased immunogenicity is also seen in cells induced to produce trehalose but which have not been subjected to a drying process.

15

SUMMARY OF THE INVENTION:

The present invention thus provides a method for producing a vaccine composition, which comprises the steps of:

20 a. Treating procaryotic cells *in vitro* under conditions such that an increase of the concentration of trehalose within procaryotic cells is induced, preferably by the synthesis of trehalose within the cell;

25 b. using the induced cells containing trehalose as the immunogenic determinant in the production of a vaccine composition.

30 Preferably, the treatment of the procaryotic cells is carried out to achieve a concentration of trehalose within the cells of at least 10mM.

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Preferably, the induced cells containing the trehalose are dried prior to their use in the production of the vaccine composition, notably in the presence of a non-reducing carbohydrate such as trehalose to provide a storage stable
5 but viable immunogenic determinant for storage prior to use in a vaccine composition.

The invention also provides a vaccine composition containing an immunogenic determinant, characterised in
10 that the immunogenic determinant has been made by the method of the invention.

The invention also provides a method for immunising an animal which comprises administering a pharmaceutically
15 effective amount of a vaccine composition of the invention to an animal sufficient to elicit an immune response in the animal.

Preferably, the vaccine composition contains an adjuvant
20 for the immunogenic determinant, is put up in an aqueous carrier medium and is administered by injection.

The prokaryotic cells for use in the present invention are ones which are capable of synthesising trehalose. This
25 ability can be native or can be conferred by recombinant techniques. The ability of a prokaryotic cell to synthesise trehalose can be determined by measuring trehalose concentration as described below.

30 The term prokaryotic is used herein to denote cells that exhibit characteristics of prokaryotes, which are typically unicellular organisms, lack organelles (such as

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mitochondria, chloroplasts, and Golgi apparatus), lack a cytoskeleton and lack a discrete nucleus. Examples of procaryotic cells for present use include bacteria, such as eubacteria, cyanobacteria and prochlorophytes; 5 archaebacteria; and other microorganisms such as rickettsias, mycoplasmas, spiroplasmas, and chlamydiae. Preferred procaryotic cells for present use are bacteria.

In general, any procaryotic cell or mixture of cells, 10 particularly bacteria, containing trehalose synthase genes should be capable of synthesising trehalose. Bacteria have two genes involved in trehalose synthesis (i.e. T- Phosphate synthase and T-P phosphatase), whereas yeasts have at least three genes and combinations of these genes 15 may be used to enable trehalose synthesis. Examples of bacteria that contain the trehalose synthase gene include, but are not limited to, Enterobacteriaceae, such as *Salmonella* and *Escherichia* (e.g., *S. typhimurium* and *E.coli*); halophilic and halotolerant bacteria, such as 20 *Ectothriphospora* (e.g., *E. halochloris*); micrococco- caceae, such as *Micrococcus* (e.g., *M. luteus*); *Rhizobium* species such as *R. japonicum* and *R. leguminosarum* bv *phaseoli*; *Cyanobacteria*; *Mycobacteria* species such as *M. tuberculosis*, *M. bovis*, and *M. smegmatis*.

25

Procaryotic cells can be induced to synthesise trehalose by culturing the cells in vitro under stressful conditions, e.g., osmotic shock, heat or oxygen limitation (shock), carbon/nitrogen starvation, or any combination of 30 the above. Suitable conditions include those heat shock and other conditions described, for example, in PCT applications Nos. GB94/01556 and GB97/03375.

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Alternatively, use of inhibitors, such as validomycin, of enzyme(s) such as trehalase involved in trehalose degradation may also result in an increase of trehalose concentration within the cells. Alternatively, the 5 genetic structure of the procaryotic organism may be modified to remove or inhibit that portion of the genetic structure which inhibits or restricts the synthesis of trehalose within the cell so that the cells constitutively synthesize trehalose as they are cultured without the need 10 to apply external stimuli. Such genetic modification can be achieved using any suitable technique. For convenience, the invention will be described hereinafter in terms of the use of external stimuli to induce the production of trehalose within the cell, rather than the 15 use of a procaryotic cell which has had its genetic structure modified.

The term osmotic shock is used herein to denote that the solute concentration in the growth medium within which the 20 cells are cultivated is above the level at which a cell exists and/or grows in its native environment. The solute may be a mixture of salts and the concentration is typically from 0.2 to 0.5 Mols above the level at which the cell is normally cultivated.

25 We believe that induction of trehalose sythesis under stressful conditions may also induce synthesis or accumulation of other molecules that may be beneficial for preservation, such as betaine and chaperonins or which 30 enhance the vaccine action of the induced cells.

For bacteria, particularly *Escherichia*, trehalose

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synthesis is preferably induced by growing the cell(s) in conditions of high osmolarity, i.e., salt concentrations sufficient to stimulate trehalose production. To induce trehalose synthesis by osmotic shock, the total
5 concentration of salt(s) in the medium should be at least about 0.2M, preferably at least about 0.4M, more preferably at least about 0.5M. The total concentration of salt(s) should not exceed 0.6M, since above this level trehalose synthesis declines in *E.coli*. The salt
10 concentrations correspond to osmolarities of at least about 350 mOsmoles to about 1.5 Osmoles, preferably at least about 400 mOsmoles to 1 Osmole, most preferably 250 mOsmoles to 500 mOsmoles. Generally, a minimum osmolarity of about 200 mOsmoles is required as this will usually
15 provide a higher concentration of solute than that under which the cells are usually cultivated.

The necessary solute can be provided by the use of a single salt, for example, 200mM NaCl KCl and/or CaCl₂.
20 (NH₄)₂SO₄ may also be used, however only about one half of the amount of trehalose is produced compared to that produced in the presence of KCl, NaCl and/or CaCl₂. A mixture of salts can also be used. In addition, when used to increase the osmolarity of the medium, a non-penetrant
25 solute such as sorbitol and/or glucose can contribute to the stimulation of trehalose synthesis.

The salt concentration (i.e., osmolarity) required to stimulate and/or induce trehalose sythesis will depend
30 upon the genus, species, and/or strain of the procaryotic cell used. Preferably, cell(s) are grown in a minimal medium containing solutes and commercially available

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minimal media can be supplemented with desired salts and/or other solutes. The use of a minimal medium is not essential and defined media can also be used. The time required to initiate and achieve the desired level of 5 trehalose concentration within the cells will vary depending on the level of osmolarity as well as the genus, species and/or strain of prokaryotic cell used. Trehalose synthesis will generally begin within an hour of placing cells in conditions designed to stimulate trehalose 10 production. Generally, in *E.coli* the synthesis of trehalose reaches a maximum at about 15-20 hours.

Synthesis of trehalose may also be stimulated using recombinant methods which are well known in the art. For 15 instance, prokaryotic cells can be transfected with a DNA plasmid comprising a DNA sequence encoding the appropriate trehalose synthase gene. The gene in turn is operatively linked to a suitable promoter, which can be constitutive or inducible. Suitable recombinant techniques are 20 described in, for example, Molecular Cloning: A Laboratory Manual, second edition (Sambrook et al., 1989).

The concentration of trehalose synthesised within the prokaryotic cells can be measured using any suitable assay 25 technique, for example by high pressure liquid chromatography (HPLC), coupled with electro-chemical detection and glucose assay (Trinder assay using trehalase) for quantitative enzymatic determination of trehalose.

30

Thin layer chromatography can be used as a qualitative method for the separation of different carbohydrates.

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Refractive index detection provides another means of detecting sugars quantitatively.

In measuring trehalose by HPLC, cells are disrupted and 5 trehalose preferentially solubilized in 70% ethanol, followed by removing triglycerides by chloroform extraction. Trehalose concentration is determined by multiplying trehalose concentration (as determined by a standard curve) by the fraction of final volume of 10 supernatant divided by pellet volume. A more detailed description of this assay is provided in Example 1.

Preferably, the synthesis is carried out to provide a concentration of trehalose within the cells of at least 15 about 10mM, for example at least about 30mM, preferably at least about 50mM, notably at least about 100mM.

Thus, in a preferred aspect the invention includes culturing the procaryotic cells under conditions that 20 stimulate intracellular production of trehalose, wherein intracellular concentration of trehalose reaches at least about 10mM, preferably at least about 30mM, more preferably at least about 50mM, notably at least about 100mM. It is particularly preferred that the 25 concentration be at least about 150mM.

The time required for stimulating trehalose synthesis depends, inter alia, on the nature of the procaryotic cells (including genus, species, and/or strain) and the 30 conditions under which trehalose induction occurs (i.e., whether by osmotic shock, oxygen deprivation, etc.). For trehalose induction by osmotic shock, the time required

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for maximum concentration of trehalose in turn depends on the degree of osmolarity as well as the particular salts used. The optimum conditions for trehalose synthesis can readily be determined by simple trial and errors tests.

5

The cultivated procaryotic cells containing the intracellular trehalose may then be frozen for storage before use as a vaccine. Alternatively storage of the vaccine can be effected by culturing the procaryotic cells 10 under conditions that increase trehalose concentration to a level effective to increase storage stability, mixing the cells with a drying solution which contains a stabilising agent, and drying the cells under conditions such that a glass is produced having less than about 5% 15 residual moisture. If a killed vaccine rather than a live vaccine is required, the cells may be killed by any suitable method, for example chemical fixation and radiation prior to processing for storage. Though the procaryotic cells may be used as the sole immunogenic 20 determinant active ingredient in the vaccine, an adjuvant may be added in an amount sufficient to enhance the immune response to the procaryotic vaccine. The adjuvant can be added to the procaryotic cells before drying, for example, cholera B toxin sub-unit can be dried simultaneously with 25 V. cholera. Alternatively the adjuvant may be obtained and dried separately, and reconstituted along with the procaryotic cells.

Suitable adjuvants include, but are not limited to, 30 aluminium hydroxide, alum, QS-21 (U.S. Pat. No 5,057,540), DHEA (U.S. Pats. Nos. 5,407,684 and 5,077,284) and its derivatives (including salts) and precursors (e.g., DHEA-

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S), beta-2 microglobulin (WO 91/16924), muramyl dipeptides, muramyl tripeptides (U.S. Pat. No. 5,171,568), monophosphoryl lipid A (U.S. Pat. No. 4,436,728; WO 92/16231) and its derivatives (e.g., DetoxTM), and BCG (U.S. Pat. No. 4,726,947). Other suitable adjuvants include aluminium salts, squalene mixtures (SAF-1), muramyl peptide, saponin derivatives, mycobacterium wall preparations, mycolic acid derivatives, non-ionic block copolymer surfactants, Quil A, cholera toxin B sub-unit, polyphosphazene and derivatives, and immunostimulating complexes (ISCOMs) such as those described by Takahashi et al. (1990) *Nature* 344:873-875. The choice of an adjuvant will depend in part on the stability of the vaccine in the presence of the adjuvant, the route of administration, and the regulatory acceptability of the adjuvant, particularly when intended for human use. For instance, alum is approved by the United States Food and Drug Administration (FDA) for use as an adjuvant in humans.

The invention also provides a method for treating an animal with a vaccine of the invention by administering a pharmaceutically acceptable quantity of the vaccine of the invention, optionally in combination with an adjuvant, sufficient to elicit an immune response in the animal. The animal is typically a human. However, the invention can also be applied to the treatment of other mammals such as horses, cattle, goats, sheep or swine, and to the treatment of birds, notably poultry such as chicken or turkeys.

30

The vaccine compositions of the present invention may be administered by any suitable means, such as orally, by

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inhalation, transdermally or by injection and in any suitable carrier medium. However, it is preferred to administer the vaccine as an aqueous composition by injection using any suitable needle or needle-less
5 technique.

The vaccines of the invention may contain any suitable concentration of the induced prokaryotic cells. We prefer that the cells are administered at doses in the range of
10 10-600 µg, preferably 10-100 µg, most preferably 25 µg, per Kg of body weight of the animal being treated. It will be appreciated that the vaccine of the invention may be applied as an initial treatment followed by one or more subsequent treatments at the same or a different dosage
15 rate at an interval of from 1 to 26 weeks between each treatment to provide prolonged immunisation against the pathogen.

20

The following examples are provided to illustrate but not limit the invention.

Example 1: Induction of trehalose in *E.coli* by osmotic
25 shock:

E.coli (NCIMB strain 9484) was cultured in Evans medium (pH 7.0) containing 5mM ammonium chloride. After overnight incubation at 37°C in the initial Evans medium,
30 a 4ml culture of *E.coli* grown in Evans medium under nitrogen limitation was used to inoculate a 200ml culture of Evans medium modified to induce osmotic shock by

- 13 -

increasing the salt concentration (KCl) to 0.5M.

Trehalose concentration was measured by high pressure liquid chromatography (HPLC) analysis and significant 5 increases in trehalose concentrations were observed at 15-17 hours after initiation of osmotic shock, with values peaking at less than 20 hours.

Example 2: Induction of trehalose synthesis in *Salmonella*:

10

Salmonella typhimurium (1344) was grown overnight at 37°C in M9 (minimal) medium with and without 0.5M NaCl. Cells were harvested by centrifugation and analysed for trehalose concentration by HPLC analysis as described in 15 Example 1. Growth in high salt medium showed at 4 to 5 fold induction of trehalose synthesis as compared to the low salt medium.

20

Example 3: Drying of prokaryotic cells after induction of trehalose synthesis:

E.coli and *Salmonella typhimurium* were grown overnight at 25 37°C in M9 (minimal) medium with and without 0.5M NaCl and trehalose synthesis induced as described in examples 1 and 2. The induced bacteria were harvested by centrifugation at 10,000 rpm for 10 minutes and the cell pellets re-suspended in drying solution containing 45% trehalose, 30 0.1% cmc (sodium carboxymethyl cellulose, Blanose 7HF, Aqualon) to a typical cell density of 0.5-1.2 x 10⁹ CFU/ml. 300µl and 500µl aliquots were dispensed into 3ml

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pharmaceutical vials and dried under vacuum without freezing, overnight at ambient temperature and a vacuum pressure of 30mTorr. Alternatively, the aliquots can be freeze-dried using the following protocol: ramp at 5 2.5°C/min to an initial shelf temperature of -40°C; primary drying was performed at a vacuum pressure of 30mT at -40°C and held for 40 hours; for secondary drying ramp at 0.05°C/min from -40 to 30°C and hold for 12 hours.

10 Example 4: Use of induced procaryotic cells as vaccines:

E.coli and *Salmonella typhimurium* cells were induced to synthesise trehalose as in Examples 1 and 2 and were used to immunise mice and rabbits. Titration of the bacteria 15 showed that a 100 to 1000 fold lower titre of bacteria induced for trehalose synthesis was required to produce an equivalent antibody response in the animals compared to the use of non-induced bacteria. Dried preparations were generally 2-50 fold more effective on a cell number basis 20 at eliciting protective immunity in the immunised animals than non-dried preparations.

Example 5: Use of induced procaryotic cells as vaccines; heat-induced trehalose synthesis:

25

E.coli and *Salmonella typhimurium* (strains as in examples 1 and 2) were grown overnight at 37°C in LB medium. 4ml aliquots of the stationary cultures were used to inoculate 200ml of LB medium in a 2 litre conical flask and the 30 cultures grown for 3hrs at 30°C. The log phase cultures were then raised to 40°C and grown for a further 3hrs before the bacteria were harvested by centrifugation at

- 15 -

10,000 rpm for 10 minutes. A similar protocol was used for the growth and induction of *Mycobacterium Bovis* and *Vaccae* (NCTC 11659) which were grown for 2 days in Sauton's medium before dilution to obtain log phase cultures for heat-induction. Cell pellets were re-suspended in lysis solution containing 0.5% Tween and the trehalose concentration was measured by high pressure liquid chromatography (HPLC) analysis. Typically 3-5 fold increases in trehalose concentrations were observed as compared to cells grown at 30°C alone.

Bacterial cells induced to synthesise trehalose as described above were killed by repeated freeze-thaw cycles and used to immunise rabbits. Antibody titres in the immunised animals were assayed by 10-fold serial dilutions using a dot-blot assay on total cell lysates prepared as described for trehalose analysis above. Animals vaccinated with induced bacteria showed a 10 to 100 fold higher antibody titre than those immunised with non-induced bacteria.

CLAIMS

1. A method for producing a vaccine composition containing an immunogenic determinant as the active ingredient, characterised in that the method comprises the steps of:
 - a. treating prokaryotic cells under conditions such that an increase of the concentration of trehalose within prokaryotic cells is induced;
 - b. using the induced cells containing trehalose as the immunogenic determinant in the production of a vaccine composition.
2. A method as claimed in claim 1, characterised in that the treatment of the prokaryotic cells is carried out to achieve a concentration of trehalose within the cells of at least 10mM.
3. A method as claimed in either of claims 1 or 2, characterised in that the increase in concentration of trehalose is achieved by synthesis of trehalose within the cell.
4. A method as claimed in any one of the preceding claims, characterised in that the condition causing the increase of trehalose concentration within the cells is heat, osmotic shock, suppression of degradation of trehalose, or genetically engineered constitutive synthesis of trehalose within the cells.

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- 1 5. A method as claimed in any one of the preceding
2 claims, characterised in that the induced cells
3 containing the trehalose are dried prior to
4 their use in the production of the vaccine
5 composition.
- 6
- 7 6. A method as in claim 5, characterised in that
8 the cells are dried in the absence of added
9 extra-cellular carbohydrate glassy stabilising
10 matrix.
- 11
- 12 7. A method as claimed in any one of the preceding
13 claims, characterised in that the procaryotic
14 cells are bacteria, protozoa or fungi...
- 15
- 16 8. A method as claimed in any one of the preceding
17 claims, characterised in that the procaryotic
18 cells are treated by cultivating them in a
19 medium containing one or more solutes and
20 having an osmolarity of at least 350 mOsmoles.
- 21
- 22 9. A method as claimed in claim 8, characterised
23 in that the solute is selected from a sodium,
24 potassium, calcium and / or ammonium salt.
- 25
- 26 10. A method as claimed in claim 1, characterised
27 in that the procaryotic cell has been modified
28 so as to synthesise trehalose.
- 29
- 30 11. A method as claimed in claim 1, characterised
31 in that the treatment of the cells is carried

1 out to achieve a concentration of trehalose
2 within the cells of at least 100mM.

3

4 12. A method as claimed in any one of the preceding
5 claims, characterised in that the procaryotic
6 cells containing the induced trehalose are
7 killed prior to use in the vaccine composition.

8

9 13. A method as claimed in any one of the preceding
10 claims, characterised in that the treatment of
11 the procaryotic cells is carried out *in vitro*.

12

13 14. A vaccine composition comprising an immunogenic
14 determinant, characterised in that the
15 immunogenic determinant includes a procaryotic
16 cell or cell residue which contains at least
17 10mM of trehalose within the cell.

18

19 15. A vaccine composition characterised in that it
20 contains an immunogenic determinant produced by
21 the method of any of claims 1 to 13.

22

23 16. A vaccine composition as claimed in either of
24 claims 14 or 15, characterised in that it
25 contains an adjuvant for the immunogenic
26 determinant.

27

28 17. A vaccine composition as claimed in any one of
29 claims 14 to 16, characterised in that it
30 contains an aqueous carrier.

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- 1 18. A vaccine composition as claimed in any one of
2 claims 14 to 17, characterised in that the
3 induced cells containing trehalose are dried in
4 the presence of a non-reducing carbohydrate to
5 provide a storage stable but viable immunogenic
6 determinant for storage prior to use in a
7 vaccine composition.
- 8
- 9 19. The use of a composition as claimed in any one
10 of claims 14 to 18 immunise an animal.
- 11
- 12 20. A method for treating an animal with a vaccine,
13 characterised in that a pharmaceutically
14 effective amount of a vaccine composition as
15 claimed in any one of claims 14 to 18 is
16 administered to the animal to elicit an immune
17 response in the animal.
- 18
- 19 21. A method as claimed in claim 20, characterised
20 in that the vaccine composition is administered
21 by injection.
- 22
- 23 22. A prokaryotic cell which has had its genetic
24 structure modified so as to remove or inhibit
25 that portion of the genetic structure which
26 inhibits or restricts the synthesis of
27 trehalose by the cell whereby the cell
28 constitutively synthesises trehalose within the
29 cell as it grows.

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(54) Title: TREHALOSE PRODUCING CELLS AS VACCINES

(57) Abstract: The present invention relates to methods for using prokaryotic cells which have been modified or induced to synthesise trehalose as vaccines and to vaccine compositions obtained thereby.

PATENT
Attorney Docket No. 8830-23

DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are stated below next to my name:

I believe I am the original, first, and sole inventor (if only one name is listed below) or an original, first, and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

TREHALOSE PRODUCING PROKARYOTIC CELLS AS VACCINES

the specification of which is attached hereto unless the following box is checked

was filed on August 18, 2000 as Application No. _____ or PCT Application No. PCT/GB00/03223 and amended on February 14, 2002 (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the patentability of this application in accordance with 37 CFR §1.56.

I hereby claim foreign priority benefits under 35 U.S.C. §119(a)-(d) or §365(b) of any foreign application(s) for patent or inventor's certificate, or §365(a) of any PCT international application which designated at least one country other than the United States, listed below and have also identified below any foreign application for patent or inventor's certificate or PCT International application having a filing date before that of the application on which priority is claimed:

PRIOR FOREIGN/PCT APPLICATION(S)

COUNTRY/OFFICE	APPLICATION NO.	DATE OF FILING	PRIORITY CLAIMED	
GB	9919732.9	August 19, 1999	<input checked="" type="checkbox"/> YES	NO <input type="checkbox"/>
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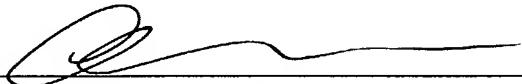
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